THE PREPARATION OF TRANS-PYRIMIDINE GLYCOLS

BY NEAR-UV IRRADIATION *

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Summary: Near-UV irradiation of Thy, Me $_2^1$, Thy, Ura, Me $_2^1$, Thy, Ura, Me $_2^1$, and Me $_2^1$, Ura in the presence of hydrogen peroxide produced corresponding cis- and trans-glycols in 2 20% yields. Similarly, Cyt, Me $_2^1$ Cyt, and Me $_2^1$ Cyt gave the corresponding glycols of Ura, Thy, and Thy, owing to facile deamination of the resultant Cyt glycols. Time sequence analysis of product-yields using $_2^1$ C-labeled pyrimidines revealed that the yields of a product are also contingent on its stability. Importantly, this approach provides a novel method for the direct synthesis of trans-glycols and for the preparation of certain radiation products in furthering our understanding of radiation chemistry and the biology of nucleic acids.

For quite some time, the possible biological implications have stimulated the study of the reactions of hydroxyl (HO·) and hydroperoxyl (HOO·) radicals with pyrimidine (Pyr) and purine derivatives (1). Such reactions have been regarded as particularly relevant to the understanding of the γ-radiation of nucleic acids in aerated aqueous solutions (2). Because of the complexities of radiation reactions, limitations by the volume and concentration of reaction solutions, and the instability of radiation products, the quantities of radiation products obtained are generally not sufficient to allow further research. This plight has impeded progress in the study of the radiation chemistry and biology of nucleic acids. However, we have taken advantage of the Milas reaction condition (3) to generate HO· and HOO· from hydrogen peroxide under the influence of near-UV light in order to prepare sufficient quantities of 5,6-dihydro-6-hydroperoxy-5-hydroxythymine (ho⁵ho₂hThy, 6-TOOH), which has been characterized as the major radiation product of Thy (4) and possibly DNA, (5) and to synthesize cis and trans Pyr glycols. Previous preparations of Pyr glycols by KMnO_4 or OsO_4 oxidation of Pyr (6,7) or by the

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treatment of corresponding Pyr bromohydrins with Ag₂O yielded only <u>cis</u> isomers (8). Such a conversion has been shown (10) to be inefficient because, for example, <u>cis</u> Ura glycol dehydrates readily form isobarbituric acid (unpublished data). For Thy glycols, the <u>trans</u> isomers which can be detected from radiolysis are in negligible amounts. Therefore, a novel approach for the <u>direct</u> syntheses of <u>trans</u> Pyr glycols in reasonable yields, as reported in this communication, becomes highly desirable.

In a typical reaction, a 10 mM aqueous solution of a Pyr [Thy, $\text{Me}_2^{1,3}$ Thy, Ura, Me^1 Ura, or $\text{Me}_2^{1,3}$ Ura] containing 40 mM of H_2O_2 was irradiated in a quartz tube with 313- or 360-nm light (11) for 24 hr. After irradiation, the excess

 $\mathrm{H_2O_2}$ and some organic peroxide formed were decomposed by treatment with 10% Pd-C. The solution was then concentrated at $\sim 35^{\circ}\mathrm{C}$ and applied to Whatman No. 3 paper or to Silica gel (Merck 60F-254) preparative tlc plates. After elution, the glycols were located by Finks reagent (12), extracted from the chromatograms with water, collected, and purified by rechromatography and/or crystallization from methanol-acetone as needed. The chromatographic and NMR spectral data are summarized in Table 1 and other data in Table 2.

Under similar conditions, Cyt, Me⁵Cyt, and Me^{4,5}Cyt gave the corresponding cis and trans glycols of Ura, Thy, and Thy, respectively. Presumably, the formation of these glycols is the result of the deamination of the respective Cyt glycols because deamination usually occurs with 5,6-hCyt derivatives (13). However, these glycols were obtained in rather low yields, possibly due to the competitive formation of corresponding Cyt N-oxides.

Time sequence analyses of product-yields with ¹⁴C-labeled compounds revealed that such yields are contingent on their stability. For instance,

Table 1. Chemical Shifts (in nnm)

		3,41	Me ₂ iny			ŝ	4.41	•	6.02	br	7.12	br	1.32	2.98	3.00)) •	l i		D 0.62	1
1. Chemical Shifts (in ppm) of NMR Spectra and R _f Values of Pyrimidine Glycols Uracil glycols (ho ^{5,6} hUra)	5,6 _{hThy})	, T, Y,	Me ₂							br	6.52	br	1.23	2.95	3.00		ı		tlc, 0.60))
	ols (ho	į	trans		I	ı	4.38		5.90		6,53	br	1.23	2.88	I	1	10.05		A 0.63	•
	ine glyc	, T	cis		1		4.45	!	5.40		6.42	br	1.25	2,89	ı	ı	10.20		paper,	
R Value	Thym	q^{Λ}	trans		ı		4.38	d,2	5.73		6.28	E	1.28		1	8.00	d,2 9.33		c 0.47	
tra and I			cis		ı		4.34	t,5	5.28		9.00	đ,5	1.28		į	8.05	d,5 9.95		paper,	
of NMR Spec		3 Ura	trans		3.86	d,5	4.63	d,5	6,50		06.9	br	ſ	2.98	3.02				B 0.91	
(mdd ur	⁶ hUra)	Me_1^1 ,	cis		4.32	d,5	4.75	d,5	5,66	br	00.0	pr	I	2.98	3.02	ı			t1c, 0.83	
Shirts (1s (5)	L Ura	trans		3,70	d,2	4.62	d, 2	6,35	a, 2	00.0	α,2	í	2,90	ı	ı	10.21 br		A 0.47	
emical	1 glyco	Me	cis		4.27	q,5	4.76	τ, ,	ν. υ.	ر م د ، م	0 7	u, J	ı	2.88	ı	1	10.18 br		paper, 0.32	
	Uraci	ຜຼ	trans		3,66	d,2	4.5I	d, 2	0.13	2, d	77.7	ţ,	ı	ı	ı	8.07	10.12		A ^e 0.34	
rable		Ura	cis	1^c	H(5) 4.20	d,2a	7 4.04	2, D	0.40	OH(6) 6.08	•	ا	!	₁₃ -	H 3 -	8.13	N(3)H 10.05 br	АРНҮ	paper, 0.22	
	*			NMR Signal $^{\mathcal{C}}$	H(5	2711	0)11	3/100	C)	0H(6		(5) #2	3	$N(1)CH_3$ –	N(3)CH ₃ -	N(1)H 8.13	N(3)H	CHROMATOGRAPHY	$\begin{array}{c} \text{System}^e \\ \text{R}_f \end{array}$	ď

 $^{\it C}{\it NMR}$ spectra were taken with a 220 MHz spectrometer. b They are also the products of Me 5 Cyt derivatives. aThey are also the products of Cyt derivatives.

^eEluents: A, 1-propanol- $^{H}_{2}$ O (10:3); B, methanol-acetone (3:1); C, 1-butanol saturated with water; D, ethyl acetate- 2 -propanol-water (75:16:9). d Coupling constants in Hz.

Table 2. Vibrational Frequencies(in cm⁻¹) of IR Spectra, Mass Spectra, and Melting Points of Pyrimidine Glycols.

								_)-18)	١٣
	3 Thy	trans		3300	ı	1148	1099sh 1111sh	1075sh 1081sh	1064		171 171	3) [‡] ((M+1	ŀ
Thymine glycols (ho ₂ ,6hThy)	$^{-}$ Me $_{2}^{1}$,	cis		3300	ı	1136	1099sh	1075sh	1053		171	((M+1)-18) ((M+1)-18) +	١٣
rcols (ho	Thy	cis trans		3250	ſ	1136	ı	1064sh	1053		174	÷(M)	155-158
ine gl_{3}	Me	cis		3250	ı	1143	1111sh	1	1053		174	(M)	² >250
Thym		trans		3425sh 3430sh	3370	1205sh	1170	1130sh	1099		142	$(M-18)^{+}$ $(M-18)^{+}$ $(M)^{+}$	145-147
	Thy	cis		3425sh	3333	1174	1114	1080	1058		142	$(M-18)^{+}$	$191-193^e$ 145-147 d >250 155-158
	3 Ura	trans		3230	í	1149	1099	1064 1064sh	1047		174	÷(₩)	٦
nUra)	$Me_2^{1,1}$	cis		3230	ı	1130	1170sh 1099	1064	1053sh		174	(M)	١٣
Uracil glycols (ho2, hUra)	Ura	cis trans		3300	ı	1176	1124	1075sh	1064		160	(M)	189-192 ^C 145-148
l glycol	Me	cis		3300	ı	1183	1136	1099	1036		160	18) [‡] (M) [‡] (189-192
Uraci]		cis trans		3356 3226sh	i	1139	1126	1111	1064		128	(M-18)	, >250
	Ura	cis	8	3356	ì	1156 1139	1139sh 1126	1087 1111	1075sh 1064		128 128	(M-18) + (M-18	229–232 ^C >250
			IR Band ^a	H-0	H-0	0=0	0=0	0-0	0	$Mass^{b}$	m/e		M.P.

 $[^]a\mathit{Spectra}$ were taken with KBr pellets.

 $[^]b$ Spectra were taken on CEC-21-110 mass spectrometer at 70 eV ionizing voltage and a source temperature of 120 o C. (Authors thank Dr. C. Fenselau and Ms. Nancy Kan for these determinations.)

 $^{^{\}mathcal{C}}_{\text{Decomposition occured during melting.}$

 $[^]d_{Product}$ appeared gummy and possibly has an m.p. close to ambiant temperature.

Melting points reported (8) are 190-191 and 195-197°C, respectively.

the yield of the relatively stable <u>cis</u> Thy glycol increased with the irradiation time; however, the yields of the relatively unstable <u>trans</u> glycols of Thy and Ura showed no further increase after reaching $^{\circ}20\%$ at 4 hr, and, as a matter of fact, a slight decrease was detected with continued irradiation. The yield of the unstable <u>cis</u> Ura glycol reached $^{\circ}20\%$ also at 4 hr; however, its gradual decrease to $^{\circ}10\%$ at 12 hr was observed. Thus, either in the collection of products or in the study of mechanisms, the stability and the photoreactivity leading to secondary reactions of particular compounds must be taken into consideration.

The above clearly indicate that sufficient quantities of certain radiation products may be prepared under specific photolytic conditions. This, in turn, permits further studies in radiation biology (14). In addition, because $\rm H_2O_2$ and $\rm O_2$ are present in biological systems, under the influence of the near-UV from sunlight, similar free-radical reactions may occur. Therefore, a detailed study of this sort has been undertaken (15) in order to gain information which may be of value to the understanding of biological near-UV effects such as aging, mutagenesis, or carcinogenesis.

REFERENCES

- Scholes, G., Weiss, J., and Wheeler, C.M. (1956) Nature, <u>178</u>, 157; Ekert, B., and Monier, R. (1959) Nature, <u>184</u>, B.A. 58; Schweibert, M., and Daniels, M. (1971) Int. J. Rad. Phys. Chem. <u>3</u>, 353; Cadet, J., and Teoule, R. (1975) Bull. Soc. Chim., 879.
- Scholes, G. (1976) in "Photochemistry and Photobiology of Nucleic Acids, Chemistry", Vol. 1, Chap. 12, Ed. S.Y. Wang, Acad. Press, New York; Cerutti, P.A., Vol. 2, Chap. 9, <u>ibid.</u>; Latarjet, R. (1972) Current Topics in Rad. Res. Quarterly, <u>8</u>, 1; Blok, J., and Loman, H. (1973) <u>ibid.</u>, <u>9</u>, 165.
- Milas, N.A., Kurz, P.F., and Anslow, W.P., Jr. (1937) J. Amer. Chem. Soc., 59, 543.
- Hahn, B.S., and Wang, S.Y. (1973) Biochem. Biophys. Res. Comm., 54, 1224;
 Teoule, R., and Cadet, J. (1974) Z. Naturforsch, 29c, 645.
- 5. Hariharan, P.V., and Cerutti, P.A. (1972) J. Mol. Biol., 66, 65.
- 6. Iida, S., and Hayatsu, H. (1970) Biochim. Biophys. Acta $\overline{213}$, 1.
- 7. Burton, K., and Riley, W.T. (1965) Biochem. J., 98, 70; Beer, M., Stern, S., Carmalt, D., and Mohlhenrich, K. (1966) Biochemistry, 5, 2283; Subbaraman, L.R., Subbaraman, J., and Behrman, E.J. (1973) J. Org. Chem. 38, 1499 and references therein.

- 8. Baudisch, O., and Davidson, D. (1925) J. Biol. Chem. 64, 233, 239.
- Latarjet, R., Ekert, B., Apelgot, S., and Reybeyrotte, N. (1961) J. Chim. Phys., <u>58</u>, 1046; Teoule, R., and Cadet, J. (1970) Bull. Soc. Chim. Fr. 925; Cadet, J., and Teoule, R. (1975) Tetrahedron <u>31</u>, 2057.
- 10. Hahn, B.S., and Wang, S.Y. (1972) J. Amer. Chem. Soc. 94, 4764.
- 11. General Electric black lamps and BLB lamps were used for the irradiation sources of 313 and 360 nm light, respectively.
- Fink, R.M., Cline, R.E., McGaughey, C., and Fink, K. (1956) Anal. Chem., 28, 4.
- Green, M., and Cohen, S.S. (1957) J. Biol. Chem. <u>228</u>, 601; Brown, D.J. (1962) "The Pyrimidines", Wiley, New York.
- 14. Thomas, H., Herriott, R.M., Hahn, B.S., and Wang, S.Y. (1976) Nature, <u>259</u>, 341; Wang, S.Y., and Parkhill, L. (1974) Radiation Res., <u>59</u>, 274; Kelley, J.E.T., Hahn, B.S., Wang, S.Y., and Merz, T. (1974) Radiation Res., <u>59</u>, 283.
- 15. Wang, S.Y., and Hahn, B.S. (1974) 2nd Annual Meeting Amer. Soc. Photobiol. Abst. WAM-D2, Vancouver, B.C., July; Hahn, B.S., and Wang, S.Y. (1974) Radiation Res. 59, 27; Cabrera-Juarez, E., and Setlow, J.K. (1977) Biochim. Biophys. Acta 475, 315.